PUBESCENOL, A WITHANOLIDE FROM PHYSALIS PUBESCENCE

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Abstract—A new with anolide, named pubescenol, and also physalin E and physalin E acetate have been isolated from *Physalis pubescence*. The structure of pubescenol is assigned as $4\alpha,7\alpha$ -dihydroxy-1-oxo- $24\alpha,25\alpha$ -epoxy-2,3-dihydrowith anolide.

INTRODUCTION

Physalis pubescence is a diploid annual of Canadian origin which is cultivated in Puerto Rico as a medicinal herb [1]. It was raised in the Botanic Farm of Andhra University, Waltair, for our chemical examination.

RESULTS AND DISCUSSION

Fresh green plants were collected, shade-dried and extracted with *n*-hexane and chloroform. The dark green *n*-hexane extract contained only plant waxes. The green chloroform extract was separated on a silica gel G column eluted with benzene—ethyl acetate mixtures to yield a new steroid, and also physalin E and its acetate. The latter compounds were identified by their NMR and mass spectra and by direct comparison with authentic samples [2, 3].

As the new steroid did not correspond with any known physalins or withanolides, it was given the trivial name pubescenol, indicating its origin from Physalis pubescence. Pubescenol (1) crystallized from acetone as colourless needles; mp 180-182°; $C_{28}H_{42}O_6$; v_{max}^{KBr} 3410 (OH), 1728 (δ -lactone), 1710 (six-membered ketone) 1050 and 1030 cm⁻¹. It contained two secondary hydroxyls (δ 3.64 br s and 5.33 d, J = 4 Hz) and yielded a diacetate (2) with pyridine and acetic anhydride at 100°; mp 201-202°; $C_{32}H_{46}O_8$; δ 1.98 s, 2.03 s (two acetoxyls). Pubescenol was characterized as a steroid resembling a withanolide, the side-chain carrying a δ -lactone system. This was supported by its ¹H NMR spectrum in which the C-18 and C-19 methyls appeared as two singlets at δ 0.66 and 0.91 and the C-21 methyl as a doublet at 0.87. There was a singlet at 1.45 (6H) accounting for two methyls and leading to the inevitable conclusion that the δ -lactone ring system in the side chain might carry an epoxide ring and the two methyls might be located on the oxirane ring system [4, 5] between C-24 and C-25.

Unequivocal support for the oxirane ring in pubescenol was obtained from its mass spectrum in which there was a prominent peak at m/z 458.2992 (rel. int. 25%), 16 amu less than the $[M]^+$ ion (474 not observed) apparently due to the loss of one oxygen atom. Such a loss of elemental oxygen is not unusual and has been noticed frequently among compounds with an oxirane system [6]. The mass

spectrum contained two more strong peaks at m/z 440.2959 (75%) and 422.2756 (25%) indicating further successive losses of 1 mol and 2 mol of water. Similarly from the pubescenol diacetate, [M] at m/z 558 (5%), the loss of one oxygen atom and 2 mol of acetic acid furnished the last mentioned ion at m/z 422 as the base peak. Furthermore, the ion at m/z 141 (45%) represented the δ -lactone system carrying an oxirane ring, while m/z 305 (4%) denoted the tetracyclic fragment without the side chain. The mass fragmentation of pubescenol and its acetate is represented in Scheme 1 in which the ions at m/z 168 (49%) and 125 (27%) are taken to represent ring A carrying one hydroxyl or acetoxyl besides the ketonic function.

The ¹H NMR spectral data of pubescenol and its acetate (Table 1) do not indicate any α,β -unsaturated ketonic system but a reduced ring A ketone as in dihydrowithanolides. The ketonic function of a six-membered ring is therefore placed at C-1 as in physalins [7] and withanolides [8]. The signal at δ 5.49 in 2 is a multiplet that can be analysed as a doublet of doublets (J = 6, 2 Hz) assignable to the 4- β H and hence the 4 α -hydroxyl is indicated to be in ring A. The alternate 3 β -position for this hydroxyl is regarded to be improbable because of its stability towards protonic reagents (see below).

The possibility of the second hydroxyl in ring C or D was eliminated by the 1H NMR resonances of the C-19 and C-18 methyls which were noticed at the same positions as in any unsubstituted steroids [9]. Pubescenol resembled Jabarosa lactone F (3) [10, 11] in its cleavage pattern in the mass spectrum. For example, the ion at m/z 228 (49%) represented the A/B/C rings as a single unit from which 2 mol of water (2 mol of acetic acid from the acetate 2) were lost. The comparative absence of the effect of the acetoxyl on the C-19 methyl resonance in the 1H NMR spectrum completely eliminated the 6 β -position for the second hydroxyl. From the multiplet at δ 4.98 for the geminal proton on the C-7 acetoxyl, the C-7 hydroxyl is regarded to be α -axial oriented.

The structure of pubescenol is thus assigned as 4α , 7α -dihydroxy-1-oxo- 24α , 25α -epoxy-2,3-dihydro-withanolide (1), which is different from a new steroid, physapubescin [5], isolated from the same plant.

Scheme 1. Mass spectral fragmentation of pubescenol (1) and its acetate (2).

Further confirmation of the presence of the 4α , 7α hydroxyls in pubescenol (1) was secured from a study of its behaviour in the presence of the protonic reagents (a) 5% H₂SO₄ in ethanol and (b) 5% H₂SO₄ in acetic acid. With the former, a trihydroxy derivative, compound 4, $C_{28}H_{42}O_6$, mp 252°, UV: nil, R_f : 0.38 (benzene-ethyl acetate, 3:7), was produced but with the latter mixture the corresponding triacetate, compound 5, C₃₄H₄₈O₉: mp 136-138°; R_f : 0.29 (benzene-ethyl acetate, 4:1) was formed. Their 1H NMR spectra revealed that one of the two hydroxyls was eliminated with the introduction of a trisubstituted olefinic bond, and also, as in physapubescin [5], the oxirane ring was cleaved to yield the $24\beta,25\alpha$ glycol. The ¹H NMR spectra (Table 1) contained no olefinic proton signals characteristic of 2-en-1-one which might be expected if the pubescenol (1) contained a 3hydroxyl group [12]. On the other hand, the spectrum of compound 4 contained a broad triplet at δ 5.60 (compound 5 had a multiplet at δ 5.52) indicating a proton on a trisubstituted olefinic bond in the molecule. Out of the three probable positions, C-4/C-5 or C-5/C-6 can be readily eliminated as these are known to cause a downfield shift of the C-19 methyl signal by ca 0.4 ppm [12]. Since it was noticed at δ 1.010 (s) in compound 4 and 1.07 (s) in compound B without a prominent shift from 0.91 in the parent compound, the double bond was therefore placed at C-7/C-8 and the olefinic proton at C-7. This olefinic bond could have arisen by the proton catalysed transdiaxial elimination of the 7α -hydroxyl in pubescenol (1). The 4α -hydroxyl was thus left intact and now the 4β -H appeared at 3.950 (m) in compound 4 and at 4.90 (m) in its acetate (compound 5).

Assignment	Pubescenol (1)*	Pubescenol acetate (2)*	Compound 4†	Compound 5
H-18	0.68	0.68 s	0.718 s	0.704 s
H-21	0.84 d (J = 6 Hz)	0.86 d (J = 6 Hz)	0.997 d (J = 6 Hz)	0.987 d (J = 6 Hz)
H-19	0.91 s	0.91 s	1.010 s	1.087 s
H-27 H-28	1.45 s	1.45 s	1.509 s 1.370 s	1.509 s 1.388 s
H-22	4.38 m	4.38 m	4.392 d/t	4.384 m
Η-4β	4.28 m	5.49 m	3.950 m	4.90 m
Η-7β	3.64 m	4.98 m	_	_
H-7			5.60 t	5.521 m
OAc-4α or OH-4α	5.33 d	2.00 s		1.999 s
OH-7 or OAc-7	4.31 m	1.98 s		
OAc-25				2.030 s
OAc-24	-	_		2.036 s

Table 1. ¹H NMR spectral data for pubescenol (1), its acetate (2) and compounds 4 and 5 produced by dehydration

In compound 4, the C-27 and C-28 methyl groups appeared at δ 1.509 and 1.370, respectively (and in 5 at 1.509 and 1.388) suggesting that they each carry a hydroxyl or acetate due to the scission of the oxirane ring in the lactone ring, as observed in physapubescin [5]

under similar circumstances. Finally, the C-22 CH proton was noted at δ 4.392 in the spectrum of 4 and at 4.384 in that of 5, values, very close to that observed in the spectrum of pubescenol (4.38) which confirms that the C-23 methylene was unchanged during this reaction.

From the foregoing results, compound 4 is assigned the structure $4\alpha,24\beta,25\alpha$ -trihydroxy-1-oxo-2,3-dihydro-7-en-22,26-olide (4) and 5 is its acetate thus providing final support for the structure of pubescenol (1).

EXPERIMENTAL

Isolation. The fresh air-dried plants of P. pubescence (500 g) were successively extracted with hexane and $CHCl_3$. The $CHCl_3$ extract was evapd and the residue (10 g) was adsorbed on silica gel (40 g) and after drying placed on the top of a column (60 × 6 cm) of silica gel G in C_6H_6 . The column was eluted with C_6H_6 -EtOAc mixtures. Sitosterol was crystallized from MeOH as colourless prisms, mp 135°, undepressed by an authentic sample. Physalin E was crystallized from MeOH as white shining plates, mp 305-307°, undepressed with an authentic sample; R_f 0.47 (C_6H_6 -EtOAc, 3:7); $[M]^+$ at m/z 544. Physalin E acetate was crystallized from MeOH as colourless shining needles, mp 278-279°, undepressed with an authentic sample.

Pubescenol (1). Crystallized from Me₂CO as colourless shining needles, mp 180–182°, R_f 0.57 (C₆H₆–EtOAc, 3:7). UV $\lambda_{\rm max}^{\rm EIOH}$ nm: 212; IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 3420 (OH), 1728 (δ -lactone), 1710 (six-membered ring ketone). (Found: C, 70.08; H, 8.89. $C_{28}H_{42}O_6$ requires: C, 70.22; H, 8.86 %) MS m/z: 458.2992 (obs.), 458.3032 (calc.) $[M-O]^+$; 440.2959 (obs.), 440.2927 (calc.) $[M-O-H_2O]^+$; 422.2756 (obs.), 422.2821 (calc.) [M-O] $-2H_2O$]⁺; 348 (rel. int. 4) [M - ring A]⁺; 330 (9) [M - ring A $-H_2O$]⁺; 329 (53); 333 (4) [M $-\delta$ -lactone -O]⁺; 305 (3) [M - side chain]⁺; 297 (22); 269 (22); 264 (5) [M - rings A/B/C]⁺; 228 (49) $[M-rings A/B/C-2H_2O]^+$; 169 (45) $[sidechain]^+$; 141 (45) $[\delta$ -lactone – O]⁺; 127 (34) $[\text{rings A} + H]^+$; 126 (27); 109 (100) [ring A – H₂O + H]⁺; 108 (12). CD (dioxan) λ_{max} nm: 295 $(\Delta t \ O)$, 265 (+0.493), 240 (-1.493), 238 (-1.454), 236 (-1.428), 233 (-1.272). Slightly negative at shorter wavelengths. ORD (dioxan): Negative Cotton effect. [M]_{288 nm} 0° (max), [M]₂₈₅ 0° (min), $[M]_{252} - 1700^{\circ}$ (max), $[M]_{248} - 1700^{\circ}$ (max), $[M]_{243}$

^{*100} MHz spectrum in DMSO-d6.

^{†270} MHz spectrum in CDCl₃.

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 -3300° (min), $[M]_{244}$ -3500° (min), $[M]_{242}$ -3100° (max), $[M]_{230}$ -4200° (min), $[M]_{225}$ $+1200^{\circ}$ (min).

Pubescenol acetate (2). This compound was prepared from 1 in pyridine–Ac₂O by heating on a steam-bath for 1 hr and crystal-lized from MeOH as colourless shining needles, mp 201–202°. IR $v_{\rm Mar}^{\rm KBr}$ cm⁻¹: 1728 (δ-lactone), 1740 (acetate), 1710 (six-membered ring ketone), 1080 and 1060. MS m/z (rel. int.): 558 [M]⁺ (5), 438 [M – 2 HOAc]⁺ (3), 422 [M – O – 2 HOAc]⁺ (100), 417 [M – δ-lactone]⁺ (3), 390 (4), 389 (9), 348 (5), 330 (10), 297 (9), 269 (20), 228 (17), 229 (50), 210 (15), 169 [side chain]⁺ (100), 168 [ring A]⁺ (49), 141 [δ-lactone + O]⁺ (100), 108 [ring A – HOAc]⁺ (100). (Found: C, 68.05; H, 8.15. C₃₂H₄₆O₈ requires: C, 68.82; H, 8.24%.]

Reaction of pubescenol with 5% H₂SO₄ in EtOH. Pubescenol (40 mg) was treated with 5% H₂SO₄-EtOH at room temp. for 10 min. The reaction mixture was coned in vacuo and after dilution extracted with CHCl₃. The solvent was removed from the product and the residue crystallized from Me₂CO. Compound 4 (R_f 0.38, EtOAc-C₆H₆, 7:3) separated as colourless cystals, mp 252°. (Found: C, 70.60; H, 9.04. C₂₈H₄₂O₆ requires: C, 70.86; H, 8.92%)

Reaction of pubescenol with 5% H₂SO₄ in glacial HOAc. Pubescenol (45 mg) was treated with cold 5% H₂SO₄-HOAc and the mixture was kept at room temp. overnight. The product was extracted with CHCl₃ and purified by chromatography over silica gel (finer than 200 mesh). Elution with C₆H₆-EtOAc (4:1) furnished compound $5(R_f 0.29; C_6H_6$ -EtOAc, 4:1) as colourless crystals, mp 136-138°. (Found: C, 67.71; H, 8.12. C₃₄H₄₈O₉ requires: C, 67.98; H, 8.05%).)

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REFERENCES

- 1. William, R. S. (1971) Lloydia 34, 165.
- Row, L. R., Sarma, N. S., Matsuura, T. and Nakashima, R. (1978) Phytochemistry 17, 1641.
- 3. Row, L. R., Reddy, K. S., Sarma, N. S., Matsuura, T. and Nakashima, R. (1978) Phytochemistry 17, 1646.
- Begley, M. J., Crombie, L., Ham, P. J. and Whiting, D. A. (1976) J. Chem. Soc. Perkin Trans. 1, 296.
- Kirson, I., Gottlieb, H. and Glotter, E. (1980) J. Chem. Res. (S) 125; J. Chem. Res. (M) 2134.
- McLafferty, F. W. (1967) Interpretation of Mass Spectra, An Introduction, p. 216. W. A. Benjamin, New York.
- Matsuura, T., Kawai, M., Nakashima, R. and Butsugen, Y. (1970) J. Chem. Soc. C 664.
- Kirson, I., Glotter, E. and Lavie, D. (1971) J. Chem. Soc. C 2032.
- 9. Kirson, I., Abraham, A., Sethi, P. D., Subramanian, S. S. and Glotter, E. (1976) Phytochemistry 16, 340.
- Tschesche, R., Baumgarth, M. and Welzel, P. (1968) Tetrahedron 24, 5169.
- Tschesche, R., Annen, K. and Welzel, P. (1972) Tetrahedron 28, 1909.
- Vande Velde, V. and Lavie, D. (1981) Phytochemistry 20, 1359.